



ALGAL FLORA OF ALCOHOL DISTILLERY EFFLUENT

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INTRODUCTION

The rate of contamination of natural water bodies increases with increased human activities in the industrial area. The wastes of such activities are finally discharged into the natural water courses resulting in the undesired eutrophication. Purdy and Butcher (1937) were among the earliest to record algal communities causing varying degree of pollution. Patrick (1948) stressed the importance of biological indicators on the assessment of water quality over the years. A number of systems have been evolved to assess the pollution of fresh waters. Brick (1963), Sladeczek (1973); Fjordingstad (1950); Dresscher and Vander Mark (1976), Sreenivasan *et al.*, (1980) have reported some fauna and flora as indicative of pollution. In the present investigation attempt were made to correlate the distribution and periodicity of algae in the desired polluted area with chemical picture of polluted water, discharged from distillery.

MATERIALS AND METHODS

The distillery is situated approximately 3 km away from Unnao city towards west of Kanpur-Lucknow railway line. The effluent of factory are discharged into an open drain which joins one of the nearby fresh water channels. Standard methods for the examination of effluent (APHA, 1976) were followed in the analytical techniques. A regular monthly sampling of effluent with simultaneous collections of algae were conducted.

Collection of Water and Algal Samples.

Spots were selected for collecting samples. Sampling was done from four or five sites in each spot. The water samples were collected at 30 days interval from the spots fixed. The samples were collected in wide mouth glass bottle (1.0 litre) and all the samples were brought to the laboratory and stored at 4^o temperature in refrigerator till the analysis was completed. The details of sampling procedure was same as described in Indian Standard methods of sampling and test for water used in industries I.S.I. New Delhi.

Samples of algae from each spots were made once a month. The samples were collected in standard

manner in bottle of about 125 cc capacity which was filled with water obtained by towing a silknet for equal distance along four side of the spots on surface and at a depth of 6-8 inch. Another bottle was filled with tips and other portion of aquatic angiosperms taken from near the surface of water. The collection being made with a little disturbance as possible. After the bottle had been left undisturbed over night the scum had settled and the clear water above was decanted off, 20 cc of material and water was left and this was preserved in 4% formalin for identification of sps.

The collected water samples were analysed for different variables by the standard methods described in Indian Standards Methods of Sampling I.S.I. New Delhi (1965). Samples were stored for further analysis after wet digestion with nitric acid and perchloric acid.

From the preserved sample algal materials were mounted on slides and examined in detail for their systematic position and periodicity.

Annual average value of important chemical parameters are furnished in Table No. 1.

RESULTS AND DISCUSSION

The occurrence and periodicity of algal samples studied are given in Table No. 2. The distribution of algae found in distillery effluent showed 14 sps. belonging to cyanophyceae, chlorophyceae and basillariophyceae.

Blue green algae : Several important publications deal with the ecological distribution of cyanophyceae. Papers of Fritsch (1907), Pearsal (1932), Yoshimura (1932), Prescott (1938), Gonzalves and Joshi (1946), Rao (1955), Singh (1960), Philipose (1960), Venkateswarlu (1969b) and Munawar (1970) have led to some controversies. Many of them emphasize the importance of light, temperature, pH, CO₂, organic matter, alkalinity, nitrates and phosphates as factors important in determining the distribution of blue-green algae.

In the present investigation the blue green algae dominate the effluent. Abundance of *Oscillatoria* sps. was frequently observed throughout the year while blooming was recorded from points where there was organic enrichment, their abundance is attributed to favourable content of oxidizable organic matter and less dissolved



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Table-1: Physico-Chemical Characteristics Distillery Effluent Variables.

| Variables | Period of Samples | | | | | | | | | | | |
|---|-------------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|------------|
| | May 2010 | June 2010 | July 2010 | Aug. 2010 | Sep. 2010 | Oct. 2010 | Nov. 2010 | Dec. 2010 | Jan. 2011 | Feb. 2011 | Mar. 2011 | April 2011 |
| Colour | Brown | Yellow | Yellow | Light | Yellow | Brown | Brown | Brown | Brown | Brown | Brown | Brown |
| Temperature | 24.5 | 22.5 | 21.9 | 24.0 | 22.5 | 22.7 | 21.2 | 20.1 | 20.9 | 21.0 | 29.2 | 28.8 |
| pH | 5.3 | 7.1 | 7.0 | 7.2 | 7.1 | 5.5 | 4.6 | 4.7 | 5.5 | 5.4 | 5.7 | 4.9 |
| Carbonates as CaCO ₃ meg/L | 4.7 | 2.3 | 1.8 | 1.0 | 0.9 | 1.2 | 5.2 | 4.5 | 4.9 | 5.2 | 5.4 | 5.3 |
| Bicarbonate as CaCO ₃ meg/L | 2.1 | 0.4 | 0.5 | 0.7 | Nil | 0.5 | 3.5 | 2.9 | 3.0 | 3.1 | 3.2 | 2.7 |
| Chloride as CaCO ₃ meg/L | | | | 47 | 59 | 50 | 525 | 563 | 549 | 546 | 403 | 521 |
| Total Alkalinity as CaCO ₃ meg/L | 1040 | 998 | 1070 | 1225 | 1360 | 1310 | 1360 | 1226 | 1553 | 1336 | 1229 | 1249 |
| Nitrite as N mg/L | Nil | Nil | Nil | Nil | Nil | Nil | Nil | Nil | Nil | 0.1 | 0.1 | Nil |
| Total Hardness as CaCO ₃ meg/L | 1260 | 691 | 784 | 860 | 847 | 919 | 2300 | 1240 | 1720 | 2100 | 1471 | 2251 |
| Alkaline hardness as CaCO ₃ meg/L | 724 | 371 | 348 | 225 | 272 | 371 | 860 | 846 | 835 | 826 | 853 | 849 |
| Non hardness as CaCO ₃ meg/L | Nil | Nil | Nil | Nil | Nil | Nil | Nil | Nil | Nil | Nil | Nil | Nil |
| Total solids in mg/L | 1547 | 772 | 906 | 1060 | 729 | 1020 | 2008 | 1920 | 1847 | 2053 | 1840 | 1912 |
| Total dissolved solids in mg/L | 1007 | 880 | 977 | 1250 | 1191 | 1421 | 1425 | 1408 | 1360 | 1235 | 1550 | 1252 |
| Total suspended solids in mg/L | 789 | 691 | 684 | 772 | 712 | 742 | 912 | 925 | 872 | 960 | 847 | 919 |
| Dissolved oxygen mg/L | 0.9 | 1.4 | 1.6 | 1.7 | 2.1 | 2.0 | Nil | Nil | Nil | Nil | Nil | Nil |
| Oxygen consumed by KN _n O ₄ in 3 hr | 1.2 | 1.6 | 1.7 | 1.7 | 1.9 | 2.1 | 1.0 | 1.1 | 1.2 | 1.2 | 1.1 | 1.1 |

oxygen. Phormidium and Aphanocapsa were more abundant in polluted zones than in clear water. Aulosira, Lyngbya, Scytonema and synecocystis were recorded in many points. Microcoleus was recorded same at one point only.

Green Algae : Waters favouring green algae are chemically distinct from those favouring blue green algae and diatoms (Pearsall 1922; Storm, 1928; Gonzalves and Joshi, 1946; Prescott, 1948; Rao 1955; Philipose, 1960; Zafer, 1964). Work of Hutchinson (1967) and Munawar (1970) have shown that even amongst the green algae volvocales, chlorococcales and desmieds have different physiological and ecological preferences. Green algae in

present study are only a few consisting of four sps with dominance of Spirogyra over Cladophora. Spirogyra was recorded throughout the year where as Oedogonium and Vaucheria were recorded at few point and a few months of a year. Volvocales are total absent.

Diatoms : Diatoms are represented by only two sps. In present study genera Navicula was found to be very bundant throughout the year. Teh abundance is attributed to favourable contents like less dissolved oxygen, oxidizable organic matter and absence of high water currents.

Table-1 : Physico-Chemical Characteristics Distillery Effluent Variables.

| Algae | Period of Samples | | | | | | | | | | | |
|------------------|-------------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|---------------|
| | May 2010 | June 2010 | July 2010 | Aug. 2010 | Sep. 2010 | Oct. 2010 | Nov. 2010 | Dec. 2010 | Jan. 2011 | Feb. 2011 | Mar. 2011 | April 2011 |
| 1. Oscillatoria | + | + | | | + | + | + | + | + | + | + | + |
| 2. Phormidium | | + | | | | + | + | + | + | + | | |
| 3. Lyngbya | | | | | | | | + | + | + | | |
| 4. Microcoleus | | | | | | + | | | | | | |
| 5. Aphanocapsa | | | | | | + | + | + | + | + | | |
| 6. Aulosira | | + | + | + | + | | | | | | | |
| 7. Scytonema | + | | | | + | + | | | | | | |
| 8. Synochocystis | + | | | | + | + | | | | | | |
| 9. Navicula | | + | + | + | | + | + | + | + | + | + | |
| 10. Pinnularia | + | | | + | + | | + | + | + | + | | + |
| 11. Spirogyra | + | | | + | + | | + | + | + | + | | + |
| 12. Vaucheria | | | | | + | | | | + | | | |
| 13. Oedogonium | + | + | | | | + | | | + | + | | |
| 14. Cladophora | + | | + | | + | | + | + | + | | + | + |

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