

# Histopathological Changes in Liver and Kidney of Heteropneustes fossilis (Bloch) on Chlorpyrifos Exposure

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### ABSTRACT

Aquatic organisms are inadvertently exposed to runoff water carrying pesticides and chemical fertilizers from agricultural land. Chlorpyrifos is one of the most commonly used organophosphate insecticides applied in agriculture for insect control. Effect of chlorpyrifos on kidney and liver in *Heteropneustes fossilis* (Bloch) was studied after exposing the fish for short term - 24h, 48h, 72h and 96h. Healthy adult fish were exposed to sub-lethal concentration 1.56 mgL<sup>-1</sup> (75% of 2.08 mgL<sup>-1</sup> 96h LC<sub>50</sub>). Histopathological changes in liver and kidney were examined at given time intervals after sacrificing fishes and tissue fixation followed by histological examination of Haematoxylin and Eosin (H/E) stained sections of tissues under light microscope. Fishes exposed to chlorpyrifos exhibited histomorphological alterations in kidney and liver while fishes in control were normal with no observation of any damage in these organs. Liver cells in exposed fishes showed necrosis and vacuolization together with random observation of denucleated cells and eosinophilic material in the cytoplasm. In kidney necrotic lesions were observed with presence of pyknotic nuclei and dilation of renal tubules. In this study it was observed that chlorpyrifos was highly toxic for the fish *Heteropneustes fossilis* and has profound adverse effect of one of the most vital organs, the liver and kidney.

Keywords: Chlorpyrifos, Heteropneustes fossilis, histopathology, kidney, liver

## INTRODUCTION

Pesticides are used for protection of crops from insect pests all over world. The sale of all categories of pesticides has increase from 850 million US dollar in 1960 to 31191 million dollar in 2005 (Zhang et al. 2011). Per capita use of pesticides has also witnessed an increase in recent years. Currently, use of pesticides in China is 13.1 kg, Japan 11.8 kg, Italy 2.5 kg and UK 3.2 kg, USA 2.5 kg and India 0.3 kg per hectare has been reported (URL 1). Many pesticides do not biodegrade as fast as introduced in the ecosystem and a large quantity persists in soil, water, and atmosphere and also in biotic system, often in the form of their metabolites. These pesticides and metabolic forms of pesticides undergo biotransformation and bio-magnification in food chain becoming more harmful for organisms at top of the food chain. Fishes, especially carnivorous and omnivorous such as H. fossilis,

are the most affected organisms in aquatic environment contaminated with pesticides and other chemicals. In India as much as 70% of the chemical formulations used for agriculture practices are believed to affect nontarget organism and to find their way to freshwater fish (Bhatnagar *et al.* 1992).

Chlorpyrifos, a broad-spectrum chlorinated organophosphate insecticide is the most used pesticide in world to control a variety of insect pests in agriculture, orchards and households. Chlorpyrifos is highly toxic to freshwater fish, aquatic invertebrates and estuarine and marine organisms (USEPA, 1989). Cholinesterase inhibition property of chlorpyrifos was observed in acute toxicity tests of fishes exposed to very low concentration (Vasudeva *et al.* 2005). Fish may be used natural bio-indicator of aquatic pollution caused by chemicals like pesticides, herbicides, fertilizers etc. (Sinaie *et al.* 2010; Joseph and Raj, 2011).

Alteration in the cellular morphology of pesticide treated fish (Abhilash and Prakasama, 2005) and their physiological functions upon exposure to different pesticide concentration have been observed by Gupta and Saxena (2006). The study of fish liver is very important in the field of aquaculture induced by many problematic condition and aquatic pollution.

The study of fish liver and kidney is very useful in assessing the health of fishes exposed to pesticides. Histopathology of liver and kidney provides indication of chemical toxicity and a useful way to study the effects of exposure of toxicants on aquatic animals. Liver is considered as one of the most important organ responsible for detoxification and bioaccumulation process. Pesticides entering in body are eventually transported in the liver, bio-transformed and detoxified in the form of water soluble substances and finally excreted out (Hodgson and Goldstein, 2001). Fish liver histopathology is an indicator of chemical toxicity and a useful way to study the effects of toxins in exposed aquatic animals (Deka and Rita, 2012). Liver is also one of the organs used as reliable biomarker of toxic injury and contaminant exposure (Stentiford et al. 2003).

Kidney plays a vital role in the maintenance of homeostasis in an organism being the key to regulate of extracellular fluid volume of composition as well as acid –base balance. It is also a target of toxic chemical, which can disrupt its function and cause temporary or permanent disruption of homeostasis. Fish kidney is composed of two parts head and body. In *Heteropneustes fossilis*, head kidney is devoid of most of the functional structures like glomeruli and consists mostly of avascular lymphoid mass. The posterior trunk kidney is functional and contains many uriniferous tubules for excretion. Histopathological changes in the kidney due to exposure of various organic and inorganic chemical compounds have been reported in several studies (Banaee *et al.* 2011; Katuli *et al.* 2014 Millar 2002; Ghasemzadeh *et al.* 2015).

#### **MATERIALS AND METHODS**

Heteropneustes fossilis, a freshwater bottom dweller catfish; weight 22.5 $\pm$ 1.75 gram and length 16.53 $\pm$ 1.07 cm, were collected from local ponds in Sultanpur district U.P. India. These fishes were fed with standard fish food pellets during acclimatization. Fish were maintained in well aerated, large plastic tank containing non-chlorinated water and acclimatized under natural photoperiods (11h light and 13h dark) for 10 days. The physicochemical feature of the normal water was maintained according to the methods described by APHA, (1998); temperature - 22 $\pm$ 1.5 °C, dissolved oxygen - 8.5 $\pm$ 0.41 mL<sup>-1</sup>, CaCO<sub>3</sub> - 154.66±5.37 mgL<sup>-1</sup> and pH - 7.5. Healthy acclimatized fishes were selected and transferred to 15L glassware tanks for conducting experiment. Fish were divided in control and test groups. Test group was further divided in four groups depicted as 24h, 48h, 72h and 96h, and five fishes in each were introduced for further treatment. Fishes in test group were subjected to exposure of 1.56 mgL<sup>-1</sup> (75% of 96h LC<sub>50</sub>) concentration of chlorpyrifos. Neither control nor test group fishes were given any food 24h before and after commencement of the test. Fish were sacrificed at different durations (24h, 48h, 72h and 96h) by spraying MS-222 (tricaine-S-methane sulfonate) solution and organs- liver and kidneys were extricated carefully. Isolated tissues were immediately preserved in Bouin's solution for further processing and histological analysis. Tissues were dehydrated and embedded in paraffin. Sagittal sections (5µ thickness) were cut using microtome. Slides were by double staining method using aqueous Haemotoxylin and alcoholic eosin stains. Photomicrographs of stained slides were made using light microscope and observed under 100x resolutions for analysis of cellular deformity.

## RESULTS

In the present research a variety of histological changes were observed in the liver and kidney of *Heteropneustes fossilis* after exposure to chlorpyrifos at 1.56 mgL<sup>-1</sup> (75% of 96h LC<sub>50</sub>). LC<sub>50</sub> value was predetermined using SPSS-20 computer program for Probit analysis.

#### Liver

The liver of control fish exhibited normal parenchymal texture generally formed of polygonal hepatocytes between which sinusoids are irregularly distributed. Hepatocytes were uniform morphologically and no vacuolization was observed. No apparent degenerative changes in sinusoids and blood vessels were marked (Fig. 1). Fishes in test group exposed to sub-lethal concentration showed moderate to severe histological changes. The degree of degenerative changes in tissues may be related with the increasing duration of chlorpyrifos exposed fishes. After 24h exposure (Fig. 2) vacuolization randomly observed becomes highly prominent after 72h (Fig. 3). Cellular degeneration of hepatocytes became more pronounced after 72h, showing dilation in blood vessels and enlargement of sinusoidal spaces. Liver tissue exposed for 96h (Fig. 4) exhibited extensive tissue damage characterized by hepatocyte degeneration, delamination and vacuolization.

### Kidney

In *H. fossilis*, kidney may be distinguished in two regions, the anterior head kidney and posterior body kidney. Head kidney consists of lymphoid tissues and has no known

role in excretion. The interstitial tissue is the main part hematopoietic tissue in the posterior body kidney and has functional nephrons. Each nephron consists of two parts, the glomerulus and the urinary tubules. Exposure of the Heteropneustes fossilis to chlorpyrifos for 96h at sub-lethal concentration (75% of 96h LC50 =1.56 mgL<sup>-1</sup>) affected kidney as observed in histo-morphological details. Kidneys in untreated control group fishes exhibited normal histological texture (Fig. 5). In treated group, degenerative changes in kidney were marked by enlargement of glomerulus and Bowman's capsule as well as increase in intercellular spaces after 24h (Fig. 6)



Fig. 1: Liver, Control (H/E, 100x)



Fig 3. Liver: 72h (H/E, 100x)

when compared to control. After 48h of exposure, loss of nuclear material was observed in few cells associated with cellular degeneration. There were marked atrophy, shrinkage and dilation in renal tubules at 72h (Fig. 7). The damage in renal tissues gradually spreads through the glomeruli, hematopoietic tissues, and cells of distal and collecting tubules. At 96h, highly degenerative changes in haematopoitic tissues associated with severe necrosis, cellular hypertrophy and swelling in renal tubules were observed. The interstitial tissues exhibited vacuolization along with deformed and undistinguished cellular boundaries (Fig. 8).



Fig 2. Liver: 24h (H/E, 100x)



Fig 4. Liver: 96h (H/E, 100x)





Fig 5. Kidney, Control (H/E, 100x)



Fig 7. Kidney, 72h (H/E, 100x)

**Fig 1.** Histo-microphotograph of Liver, Control Fish (100x, H/E), showing normal cytoarchitecture

Fig 2. Histo-microphotograph of Liver, 24h Chlorpyrifos exposed Fish (100x, H/E), showing vacuolization (V) and degeneration (D) in hepatocytes

**Fig 3.** Histo-microphotograph of Liver, 72h Chlorpyrifos exposed Fish (100x, H/E), exhibiting marked cellular degeneration (D) of hepatocytes, dilation in blood vessels (B) and enlargement of sinusoidal spaces (E)



Fig 6. Kidney, 24h (H/E, 100x)



Fig 8. Kidney, 96h (H/E, 100x)

**Fig 4.** Histo-microphotograph of Liver, 96h Chlorpyrifos exposed Fish (100x, H/E), exhibiting hepatocyte degeneration (D), delamination and vacuolization (V)

**Fig 5:** Histo-microphotograph of Kidney, Control Fish (100x, H/E): No apparent abnormal changes

**Fig 6:** Histo-microphotograph of Kidney, 24h chlorpyrifos exposed Fish (100x, H/E): Kidney showing glomerulur enlargement (E), increase in intercellular spaces (S) and loss of nuclear material (N)

**Fig 7:** Histo-microphotograph of Kidney, 72h chlorpyrifos exposed Fish (100x, H/E): Kidney tissues showing glomerular atrophy (A) and shrinkage, and tubular dilation (D).

**Fig 8:** Histo-microphotograph of Kidney, 96h chlorpyrifos exposed Fish (100x, H/E): Kidney tissues exhibiting degeneration (D), necrosis, hypertrophy and swelling in renal tubules (H), vacuolization and deformation in cellular boundaries (V).

### DISCUSSION

Contamination of aquatic ecosystem by pesticides has adverse effects on various non-target organisms, especially the fish. Numerous studies have reported chlorpyrifos toxicity causing alterations in biochemical parameters and histological damage in vital organs like liver, kidney etc. (Bernet *et al.* 1999; Ghorashi *et al.* 2013).

#### Liver

In present study, short term (96h) chlorpyrifos exposure has shown deleterious effects on liver and kidney. Histological studies are useful tool to assess sub-lethal and chronic exposure of fishes to aquatic pollutants. Liver is the primary site of detoxification and one of the most important organs affected by pesticide exposure. Most common adverse effects observed in liver due to pesticide exposure include, cellular deformity associated with nuclear hypertrophy and vacuolization, sinusoidal enlargement, necrosis and congestion of sinusoidal spaces by WBCs infiltration. The exposure effects become moderate to severe with increasing exposure duration as revealed by the results (Fig. 1-4).

Liver in chlorpyrifos exposed H. fossilis after 24h (Fig. 2) in this study exhibited randomly located vacuolization becoming highly prominent after 72h (Fig. 3) of exposure. Besides these changes, other histopathological conditions observed in liver at 96h were, cellular degeneration of hepatocytes, dilation in blood vessels, enlargement of sinusoidal spaces, extensive tissue damage characterized by hepatocyte degeneration, necrosis and delamination of cellular boundaries were also observed in exposed fishes (Fig. 4). These findings are in agreement with that of Manjunatha and Philip (2015) and Barbhuiya and Dey (2014) who reported similar affects in Heteropneustes fossilis on exposure to sub lethal concentration of 0.12  $\mu$ l/L (1/10th of LC<sub>50</sub>) chlorpyrifos. Devi and Mishra (2013) have also made similar observations in Channa punctatus after hilban (chlorpyrifos) treatment. However, different authors observed different toxicological effects in the liver of fish after exposure to different toxicants. Kaoud et al (2010) have reported congestion of central vein in fish liver. Degenerative changes in hepatocytes like necrosis,

vacuolization and delamination of cellular boundaries reported by Velisek *et al* (2009) and Abdel-Hameid (2009) are in agreement with findings of this study. In another study, Indirabai *et al* (2010) have observed same results in malathion treated *Heteropneustes fossilis* as found in this study. Rahman *et al.* (2002) also reported similar effect of diazinon in *Anabas testudineus*, *Channa punctatus* and *Barbodes gonionotus* exposed at sub-lethal concentration of the pesticide for 7 days.

### Kidney:

The kidney of fish receives the largest proportion of post branchial blood, and therefore renal lesions might be expected to be good indicators of environmental pollution. It has been found that the sub-lethal doses of QP induced alterations in the structure of kidney of silver barb. These results reveal that the histopathological changes are dose and duration dependent. The proximal tubules were the first to be affected resulting in the reduction of nuclear material in some cells. The damage in kidney tissues further advances gradually through the glomeruli, hematopoietic tissues, and cells of distal and collecting tubules in that order (Fig. 6-8). Renal tubules proliferation and dilation, necrosis of tubular epithelium and pyknotic nucleus were seen in the tissue treated with chlorpyrifos in this study (Fig. 8). These observations are in agreement with Subburaj et al. (2020) in the kidney of Oreochromis mossambicus exposed to chlorpyrifos (50%EC) for 96h. Raibeemol and Chitra (2018) have also reported reduced kidney functions and cellular disorganization in a freshwater fish, Pseudetroplus maculatus when exposed to chlorpyrifos. In agreement to the results of this study Banik et al. (2016) have reported pyknosis, necrosis of tubular and hematopoietic cells in G. giuris after diazinon exposure. Ghasemzadeh et al. (2015) have reported injury the kidney of spotted scat fish, Scatophagus argus when exposed to diazinon for short-term (96h). Diazinon induced damage in kidney has also been reported by Khosravi et al. (2014). Similar results were observed in kidney of Gambusia affinis exposed to chlorpyrifos (Sabiha, K. and Neela, S. 2013). Kidney lesions consisting of dilation of Bowman's space has been reported by Wannee et al. (2002) which in agreement to our observation.

# CONCLUSION

The histopatholgical alterations observed in liver and kidney are due to acute toxic effect of chlorpyrifos in *Heteropneustes fossilis*. The study is useful in understanding harmful effects of pesticides of aquatic life.

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# REFERENCES

- Abdel-Hameid, N.A.H. (2009). A protective effect of calcium carbonate against arsenic toxicity of the Nile catfish, *Clarias gariepinus. Turk. J. Fish. Aquat. Sci.* 9: 191-200.
- Abhilash, R. and Prakasama, V.R., (2005). Toxic, physiomorphological and behavioural responces *Oreochromis mossambicus* exposed to commercial grade endosulfan. *Ecology*. 52(2), 234-238.
- Banaee, M., Mirvaghefi, A.R., Mojazi Amiri, B., Rafiee, G.R. and Nematdost, B. (2011). Hematological and histopathological effects of diazinon poisoning in common carp (*Cyprinus carpio*). J. Fish Iran J. Nat. Res. 64:1–13.
- Barbhuiya H.S. and Dey M. (2014). Histopathological Changes in Liver Tissue of *Heteropneustes fossilis* Exposed Chlorpyrifos (20% EC). *Indian J. App. Res.* 4(7), 237-240.
- Bernet D., Schmidt H., Meier W., Burkhardt-Holm P., Wahli T. (1999). Histopathology in fish: proposal for a protocol to assess aquatic pollution. J. Fish Disease. 22:25-34. (https://doi.org/10.1046/j.1365-2761.1999.00134.x )
- Bhatnagar, D., Ehrlich, K.C., and Cleveland, T.E. (1992).
  "Oxidation-reduction reactions in biosynthesis of secondary metabolites," In *Handbook of Applied Mycology: Mycotoxins in Ecological Systems*, Eds. D. Bhatnagar, E. B. Lillehoj, and D. K. Arrora (New York, NY: Marcel Dekker), 255–286.
- Deka, S. and Rita, M. (2012). A study on the effect of organophosphorous pesticide malathion on hepatorenal and reproductive organs of *Heteropneustes fossilis* (Bloch). *The Science Probe*. 1(1): 1-13.
- Devi, Y. and Mishra, A. (2013). Histopathological alterations in gill and liver anotomy of fresh water, air breathing fish *Channa punctatus* after pesticide hilban (chlorpyrifos) treatment. *Adv. Biores.* 4 (2): 57-62.
- Ghasemzadeh, J., Sinaei, M., Bolouki, M., (2015). Biochemical and histological changes in fish, spotted scat (*Scatophagus argus*) exposed to diazinon. *Bull. Environ. Contam. Toxicol.* 94:164–170. DOI 10.1007/s00128-014-1454-8.
- Ghorashi S., Shajeei H., Vaezi G., Shamoushaki M.M.N. and Babakhsni A. (2013). Histopathological studies on Kidneys and Gills of *Onchorhynchus mykiss* exposed to sublethal concentration of ethylene diamine tetraacetic acid (EDTA). *Global Veterinaria*, 10(2): 121-

127.

- Gupta, P. and Saxena, P.G (2006). Biochemical and haematological studies in freshwater fish *Channa punctatus* exposed to synthetic pyrethroids. *Poll. Res.* 25(3): 499-502.
- Hodgson, E. and Goldstein, J.A. (2001). Metabolism of toxicants: Phase I reactions and pharmacogenetics.
  In: Hodgson E, Smart RC, editors. *Introduction to Biochemical Toxicology*. New York: Wiley; 2001. pp. 67–113.
- Indirabai, W.P.S., Tharani, G. and Seetha, P. (2010). Impact of sublethal concentration of Endosulfan on biochemicals and histology of organ tissues of freshwater fish, *Labeo rohita* (Hamilton, 1822). *The Bioscan.* 5(2): 215-218.
- Joseph, B. and Raj, S.J. (2011). Impact of pesticide toxicity on selected biomarkers in fishes. *Int. J. Zoolog. Res.* 7(2): 212-222.
- Kaoud, H.A. and El-Dahshan, A.R. (2010. Bioaccumulation and histopathological alterations of the heavy metals in *Oreochromis niloticus* fish. *Nature and Science*. 8(4): 147-156.
- Katuli K.K., Amiri B.M., Massarsky, A., Yelghi, S., Ghasemzadeh, J. (2014). Impact of a shortterm diazinon exposure on the osmoregulation potentiality of Caspian roach (*Rutilus rutilus*) fingerlings. *Chemosphere*. 108: 396–404. http:// dx.doi.org/10.1016/j.chemosphere.2014.02.038.
- Manjunatha B. and Philip G.H. (2015). Histopathological alterations in liver anatomy after exposure to chlorpyrifos in Zebrafish (*Danio rerio*). *Der Pharmacia Lettre*, 7 (7):191-197.
- Miller, G.G., Sweet, L.I., Adams, J.V., Omann, G.M., Passino-Reader, D.R. and Meier, P.G. (2002). *In vitro* toxicity and interactions of environmental contaminants (Arochlor1254andmercury)andimmunomodulatory agents (lipopolysaccharide and cortisol) on thymocytes from lake trout (*Salvelinus namaycush*). *Fish and Shellfish Immunology*. 13:11-26.
- Rahman, M.Z., Hossain, Z., Mollah, M.F.A. and Ahmed, G.U. (2002). Effect of Diazinon 60 EC on Anabas testudineus, Channa punctatus and Barbodes gonionotus. Naga, The ICLARM Quarterly. 25(2):8-12.
- Raibeemol, K.P., Chitra, K.C. (2018). Effects of chlorpyrifos as inducer for oxidative stress in liver, kidney and spleen of freshwater fish, *Pseudetroplus maculatus* (Bloch, 1795). *Research & Reviews: A Journal of Toxicology*. 8(1): 20-29.
- Sabiha, K. and Neela, S. (2013). Histopathological alterations in the kidney of *Gambusia affinis* after exposure to chlorpyrifos. International Jouranal of Chemistry and *Pharmaceut. Sci.* 1: 122-127.

- Sinaie M., Darvish B.K., Ghorbanpour, M., Najafzadeh, H., Shekari, M., Haghparast, S. (2010). Metallothionein biosynthesis as a detoxification mechanism in mercury exposure in fish, spotted scat (*Scatophagus* argus). Fish. Physiol. Biochem. 36(4):1235-1242.
- Stentiford, G.D., Longshow, M. Lyons, B.P. Jones, G.J; Green, M. and Feist, S.W. (2003). Histological biomarkers in estuarine fish for assessment of biological effects of contaminants. *Mar. Environ. Res.* 55(2):137-159.
- Subburaj, A., Jawahar, N., Srinivasan, A. and Ahilan, B. (2020). Effects of acute toxicity of chlorpyrifos (ec 50%) and associated histological alterations in gills, liver and kidney of Mozambique tilapia, *Oreochromis mossambicus* (Peters, 1852). *Ind. J. Anim. Res.* 54(12): 1497-1504.
- URL1: https://www.worldometers.info/food-agriculture/ pesticides-by-country/
- USEPA, (1989). Risk assessment: guidance for superfund. In: Human Health Evaluation Manual (Part A), Interim Final, vol 1. Office of Emergency and Remedial

Response, U.S. Environmental Protection Agency, Washington DC.

- Vasudeva, R.Y. and Charkrabarti, R. (2005). Stimulation of immunity in Indian major carp *Catla catla* with herbal feed ingredients. *Fish and Shellfish Immunol*. 18(4): 327-334.
- Velisek, J., Svobodova, Z., and Piackova, V. (2009). Effects of acute exposure to bifenthrin on some haematological, biochemical and histopathological parametersof rainbow trout (*Oncorhynchus mykiss*). *Veterinarni Medicina*. 54(3): 131–137.
- Wannee, J., Upatham, E.S., Kruatrachne, M., Sanhaphong, S., Vichasri-Grams, S. and Pkethitiyook, P. (2002). Histopathological effects of round up, a glyphosate herbicide on Nile tilapia *Oreochromis niloticus*. *Science Asia*. 28(2): 121-127.
- Zhang, W., Fubin, J., Jianfeng, O. (2011). Global pesticide consumption and pollution: with China as a focus. *Proceedings of the International Academy of Ecology and Environmental Sciences*. 1(2):125-144.