

The Scientific Temper VOL-IX, NO.1&2; JANUARY-JULY, 2018 ISSN 0976 8653, E ISSN 2231 6396 UGC SR NO 2535; JR NO. 47226 e-mail:letmepublish@rediffmail.com Web: www.scientifictemper.com

IMPACTS OF MALATHION ON BIO-CHEMICAL CHANGES IN FRESHWATER FISH CHANNA PUNCTATUS UNDER LABORATORY CONDITIONS

Nagendra Kumar Yadav Ex-Research Scholar, JaiPrakash University, Chapra (Bihar) Email ID:nagendrakumaryadav47@gmail.com

ABSTRACT

The fresh water fish Channa punctatus was exposed to malathion in the laboratory to study its toxicity. The acute toxicity tests were conducted during certain intervals in various concentrations of malathion. The physical and chemical analyses of water were carried out by following APHA methods. The lethal and sub-lethal concentration of malathion were found to be LC100 (25 mg/L) and LC0 (5 mg/L), respectively. The antioxidant enzyme activity in the liver, muscle and gill, respectively increased during the accumulation of malathion, whereas it decreased respectively during depuration period. The effects of malathion resulted in the gradual decrease of nucleic acids, protein, free amino acids (FAA) and glycogen. During recovery period, the levels of biochemical components progressively increased indicating a probable recovery from the disruption of internal organ. Hence, the pesticide intoxication has made defective consequences in the normal metabolic pathways which led increasing the rate of mortality in fish population.

Key words: Labeo rohita, Malathion, Protein, Nucleic acids and Antioxidant enzymes

INTRODUCTION

The malathion contamination of ponds is a potential problem for aquaculture in tropical countries. The pesticide, on reaching to aquatic systems, greatly influences the non target organisms such as fish and birds. Histological studies on fish have revealed that various toxicants have produced pathological changes in the tissues such as macrobiotic changes in the liver, tubular damage of kidneys, gill and lamellar abnormalities (Ramalingam, 2000). Due to growth of agriculture in and around fresh water bodies the pesticides are used abundantly during the cultivation season and found their way into water bodies.

The degree of toxicity produced by the poisonous substance is dose independent upon environmental conditions such as temperature, pH, oxygen content and presence of residue molecules (Singh and Mishra, 2009). It is well known that protein, carbohydrates and lipid play a major role as energy precursors in fish under stress conditions. Enzymes play significant role in food utilization and metabolism. The proteolytic enzymes participate in the breakdown of protein molecules into amino acids and these amino acids are in turn oxidized to give energy for body function (Saravanan et *al.*, 2000). Pollutants can produce metabolic changes at cellular levels by a way of influencing enzyme systems.

The present study has been made to investigate the biochemical changes followed by mortality in the fresh water fish *Channa punctatus* induced by sub lethal dosages of the pesticide.

MATERIALS AND METHODS

The collected Fishes were fed daily and acclimatized in laboratory for 30 days. The physical and chemical analyses of the water were carried out (APHA, 2005). Fish were divided into seven groups (each containing 10 fish) where six were experimental and one group as control. Acute toxicity study was carried out using the standard guidelines to determine the lethal (LC100), median (LC50) and safe sub lethal (LC0) levels of malathion in various concentrations (5, 10, 15, 20, 25 & 30 mg/L). The mortality of fish (%) was assessed during the interval of 24, 48, 72 and 96 hours. The 1/3rd of median lethal concentration (5 mg/L) was taken to study the effect of malathion on the biochemical constituents and detoxifying ability of fish.

The water was renewed freshly every day to produce constant effect of malathion on fish. At the end of 15 days exposure, the tissues such as liver, muscle and gill were collected by dissecting the animal and stored at - 20°C for biochemical parameters studies. The remaining fish released into freshwater for 15 days to know the detoxifying ability of the fish. At the end of 30 days, tissues were collected again and one gram of muscle, liver and gill samples were suspended in 5mL of 0.1 M phosphate buffer of pH=7 and homogenized. These homogenates were stored for further studies at -20°C. The Catalase activity assay was performed according to Beaumont et al (1990) by following the H₂O₂ dismutation at 240 nm in a reaction mixture composed of 0.1 M phosphate buffer, pH=7, 50-100 mg protein and 18 mM H₂O₂. GST activity was measured at 37°C using 1 mM l-chloro-2,4 dinitrobenzene (CDNB) as substrate.

The activity of acid phosphatase and alkaline phosphatase were assayed with the method of TennisWood et *al* (1976). Proteins levels were estimated by the method of Lowry et *al* (1951) using bovine serum albumine as standard. Homogenates 2 ml (w/v) cold distilled water was prepared in 30% TCA; values are expressed as mg/100 mg wet wt of tissue. Free amino acids (FAA) were estimated using the ninhydrin method (Moore and Stein, 1954). .FAA was expressed as mg/100 mg wet wt of the tissue.

The values were expressed as mean \pm SEM. Statistical analysis was performed by oneway analysis of variance (ANOVA) followed by LSD tests using the computer package SPSS 18.0 v and the significance of difference was set up at (p< 0.05).

RESULTS AND OBSERVATIONS

The percentage of mortality of *Labeo rohita* exposed to malathion in 5, 10, 15, 20, 25 and 30 mg/L for 24h, 48h, 72h and 96h was assessed (Table 1).

The Scientific Temper Vol-IX, 2018

Table 1	l:	Mortality of Channa punctatus
		exposed to Malathion.

S	Concen-	Expo	Exposure period (hours)					
No.	tration							
	(mg/l)	24	48	72	96			
1	5	N	Ν	Ν	Ν	15 mg/l		
2	10	Ν	10%	10%	10%			
3	15	10%	20%	30%	50%			
4	20	20%	50%	70%	90%			
5	25	60%	100%	Ν	Ν			
6	30	10%	Ν	Ν	Ν			

The median lethal concentration was observed as 15mg/L since it is caused 50% mortality in 96 h using the "Maximum likelihood method" (Finney, 1971). 1/3rd of median lethal concentration (5 mg/L) was taken to study the effect of malathion on the biochemical constituents and detoxifying ability of fish.

Table 2: Anti-oxidant enzyme activity in the tissues of Channa punctatus during accumulation
and de-purination periods

Accumulation study	/ (umole of Phenol liberated/min/100 r	ng Protein)
accumulation study	(µmore of 1 nenor moer area, mm, 100 h	ng i i otemi

S.	Anti-Oxidant enzyme	Liver		Muscle		Gill			
No.		Control	Day 15	Control	Day 15	Control	Day 15		
1	Catalase	13.6±1.13	42.8±2.1	6.8±0.10	16.2±0.54	6.7±0.13	23.4±0.15		
2	Glutathione 5-transferase	19.5±1.12	269.8 ± 0.14	92.9±0.12	$142.9{\pm}1.04$	119±0.52	214.8±0.70		
	Accumulation study (µmole of Phenol liberated/min/100 mg Protein)								
1	Catalase	38.03 ± 1.03	16±1.42	10.3 ± 0.72	7.9±0.23	21.9±0.84	9.7±0.64		
2	Glutathione 5-transferase	251.9±1.40	218.8±1.10	134.9 ± 0.16	107.8±0.32	17.9 ± 0.27	158.2 ± 0.44		

The activity of antioxidant enzymes in the liver, muscle and gill of *Labeo rohita* exposed to LC0 concentration of 5 mg/L malathion during accumulation were observed as showed in Table 2.

Depletion on biochemical parameters like Protein, Glycogen and Free amino acid were evaluated during various periods of exposure (Table 3).

Table 3 : Sub-lethal effects of Malathion on Protein, Glycogen and Free amino acid in the	tissues
of Channa punctatus.	

_				-			
S	Organs	Biochemical	Control	Sub-lethal Concentrations			
No.		parameters		24hr	48 hr	72 hr	96 hr
1	Liver	Protein	225.3±1.42	206.68±1.20	180.2 ± 0.44	178.86±0.42	142.2 ± 0.78
		Glycogen	11.2±0.32	10.4±0.16	9.6±0.15	8.7±0.22	6.2±0.16
		FAA	28.4±0.15	27.4±0.15	35.4±0.26	33.1±0.22	28.2±0.42
2	Muscle	Protein	189.3±0.3	172.4±0.42	140.24±0.26	134.8±0.12	127.3±0.14
		Glycogen	9.6±0.3	9.2±0.3	7.4±0.16	6.7±0.26	5.2±0.16
		FAA	217.4±1.13	213.4±0.52	28.4±0.52	24.4±0.10	20.4±0.24
3	Gill	Protein	198.3±0.32	162.6±0.54	125.3±0.63	96.9±0.12	86.3±0.15
		Glycogen	11.2±0.23	9.46 ± 0.02	9.2±0.04	6.8±0.11	5.2±0.17
		FAA	22.4±0.15	22.2 ± 0.48	21.6±0.46	20.4±0.12	19.8±0.25

Reduction on macro and micromolecules are directly proportional to the concentration of malathion and exposure periods. The values were expressed as mean \pm SEM and the significance of difference was set up at (p< 0.05).

DISCUSSIONS

The fish were seen to exhibit several behavioural responses, such as fast jerking, frequently jumping, erratic swimming, spiraling, convulsions and tendency to escape from the aquaria during study.

The Scientific Temper Vol-IX, 2018

Rao et *al* (2005) reported that abnormal changes in behavior in mosquito fish *Gambusia affinis* in response to the sub-lethal exposure to chlorpyrifos.

The fish exhibited unrest and a peculiar tumbling motion before they died. Moreover, the herbicide butachlor persists in the aquatic system for a long period of time. The liver, muscle and gill tissues showed decreased level of acid phosphatase (ACP) and Alkaline Phosphatase (ALP) activities. Shakoori et *al* (1992) have suggested the decrease (or) inhibition of ACP and ALP activities are due to increased necrosis in the tissues like hepatocytes.

The protein, glycogen and free amino acids were decreased gradually compared to control, when the period of exposure increased. The depletion of protein may also be attributed to spontaneous utilization of amino acids in various catabolic reactions inside the organism in order to combat the stress condition (Borah, 1996). Increase of total free amino acids (TFAA) is an induction of stepped up proteolysis or fixation of ammonia into keto acids resulting in amino acid synthesis. Generally, these two processes contribute to the amino acid pool (Mohapatra and Noble, 1992). The carbohydrate reduction suggests the possibility of active glycogenolysis and glycolytic pathway to provide excess energy in stress condition (Reddy et al (1993).

The present investigation shows biochemical changes due to sub lethal concentration of Malathion in total proteins, free amino acids (FAA) and glycogen in target organs and tissues significantly. Thus the pesticides intoxification has disturbed the normal functioning of cells with the resultant alterations in the fundamental biochemical mechanisms in fish. This would in turn result in the mortality of fish on chronic exposure to the pesticide.

ACKNOWLEDGEMENTS

I am thankful to Head, PG Department of Zoology, JaiPrakash University, Chapra (Bihar) to provide laboratory facility for this investigation.

REFERENCES

- APHA (2005). Standard Methods for the examination of water and waste water. Standard Methods for the examination of water and waste water. 21st Ed. Washington DC.
- Borah S and Yadav RNS (1996a). Effects of rogor (30% w/v dimethoate) on the activity of lactate dehydrogenase, acid and alkaline phosphatase in the muscle and gill of fresh water fish *Heteropneustes fossilis*. J. Environ. Biol., 17(4): 279-283.
- 3. Finney DJ (1971). Probit Analysis. 3rd Ed. Cambridge University Press, London, 330 pp.
- Lowry O.H., Oserrought, MJ and Randoll, RJ (1951) Protein measurement with the Folin phenol reagents. J. Biol. Chem., 193:265-275.
- Mohapatra BC and Noble A (1992). RNA-DNA ration as indicator of stress in fish. Com. Physiol & Ecol., 17(2): 41-47.
- Moore S and Stein WH (1954). A modified ninhydrin reagent for the photometric determination of amino acids and related compounds. J. Biol. Chem. 211: 907-913.
- Ramalingam V, Vimaladevi V, Narmadaraji R and Prabakaran P (2000). Effect of lead on haematological and biochemical parameters in freshwater fish *Cirrhina mrigala*. Pollu.Res., 19:81-84
- Rao JV, Ghousia B, Pallela R, Usman PK. and Nageswara Rao R (2005). Changes in behavior and brain acetylcholinesterase activity in mosquito fish, *Gambusia affinis* in response to the sub-lethal exposure to chlorpyrifos. Int. J. Environ. Res. Public Health., 2(3): 478-483.
- Reddy MM, Kumar VA and Reddy SNL (1993). Phenol induced metabolic alteration in the brain and muscle of fresh water fish *Channa punctatus* during sublethal toxicosis. J. Ecotoxicol. Environ. Monit., 3(1):7-1
- Saravanan TS, Aneez Mohamed M and .Harikrishnan R (2000). Studies on the chronic effects of endosulfan on blood and liver of *Orechromis mossambicus*. J. Ecol. Res. Biocon., 1:24 -27.
- Shakoori AR, Alam J, Aziz F, Aslam F and Sabir M (1992). Toxic effect of bifenthrin (Talstar) on the liver of *Gallus domestics*. J. Ecotoxicol. Environ. Monit., **21**(1): 1-11.
- Singh S and Mishra RN (2009). Occurrence of organochlorine pesticides residues in Kuano river of eastern Uttar Pradesh. J. Environ. Biol., 30:467-468.
- Tennis Wood MC, Bind E and Clark AF (1976). Phosphatases antigen dependent markers of rat' prostate. Can. J. Biochem., 54: 340-343.

http://www.scientifictemper.com/